Themed Issue: Drug-Induced Hypersensitivity Reactions

Guest Editor - Craig Svensson

Mechanisms of Drug-induced Delayed-type Hypersensitivity Reactions in the Skin

Submitted: September 1, 2005; Accepted: September 12, 2005; Published: December 9, 2005

Sanjoy Roychowdhury¹ and Craig K. Svensson¹

¹Division of Pharmaceutics, College of Pharmacy, The University of Iowa, Iowa City, IA 52242

ABSTRACT

Cutaneous drug reactions (CDRs) are the most commonly reported adverse drug reactions. These reactions can range from mildly discomforting to life threatening. CDRs can arise either from immunological or nonimmunological mechanisms, though the preponderance of evidence suggests an important role for immunological responses. Some cutaneous eruptions appear shortly after drug intake, while others are not manifested until 7 to 10 days after initiation of therapy and are consistent with delayed-type hypersensitivity. This review discusses critical steps in the initiation of delayed-type hypersensitivity reactions in the skin, which include protein haptenation, dendritic cell activation/migration and T-cell propagation. Recently, an alternative mechanism of drug presentation has been postulated that does not require bioactivation of the parent drug or antigen processing to elicit a drug-specific T-cell response. This review also discusses the role of various immune-mediators, such as cytokines, nitric oxide, and reactive oxygen species, in the development of delayed-type drug hypersensitivity reactions in skin. As keratinocytes have been shown to play a crucial role in the initiation and propagation of cutaneous immune responses, we also discuss the means by which these cells may initiate or modulate CDRs.

KEYWORDS: cutaneous drug reactions, delayed-type hypersensitivity, dendritic cells, keratinocytes, T-cells, cytokines

INTRODUCTION

Adverse drug reactions (ADRs) are very common in clinical practice.¹ ADRs are noxious or unintended reactions to a drug that is administered in standard doses by the proper route for the purpose of prophylaxis, diagnosis, or treatment of a specific disease.² For some drugs, ADRs occur in a large portion of those who receive the drug (eg., nausea associated

Corresponding Author: Craig K. Svensson, Division of Pharmaceutics, College of Pharmacy, The University of Iowa, 115 S Grand Avenue, S213 PHAR, Iowa City, IA 52242. Tel: (319) 335-8823; Fax: (319) 335-9349; E-mail: craig-svensson@uiowa.edu

with oncolytic agents). Other reactions occur in only a small portion of patients receiving the agent and it is not possible to identify in advance who will experience such reactions (eg, anaphylaxis after penicillin administration). Among various target organs, skin is one of the most frequent sites for ADRs.^{3,4} Although numerous mechanisms have been suggested for cutaneous drug reactions (CDRs), experimental evidence for the mechanism of these reactions is limited.

Many of the cutaneous reactions associated with drug therapy are believed to be immune-mediated. Such reactions can be divided into 2 main categories: immediate-type or delayed-type immune-mediated reactions⁴. These CDRs are mediated either by immunoglobulin E (IgE; immediate-type reactions) or T cells (often referred to as delayed-type hypersensitivity [DTH]).⁵ While the mechanism of IgE-mediated reactions is better understood, the genetic basis for predisposition to these reactions following drug administration (eg, penicillin) is unclear.¹ Less well understood is the mechanism by which DTH reactions occur in the skin after the administration of drugs systemically. In this review, we discuss critical steps and factors associated with DTH reactions in skin and discuss potential mechanisms by which these may be provoked after systemic drug administration.

CHARACTERIZATION OF DELAYED-TYPE HYPERSENSITIVITY REACTIONS IN SKIN

Contact Dermatitis

Allergic contact dermatitis (ACD) is one of the most studied forms of xenobiotic-induced DTH, in which topical application and elicitation of sensitization to xenobiotics is confined to the skin. Eczema and dermatitis are used synonymously to denote the polymorphous pattern of skin inflammation characterized in its acute phase by erythema, vesiculation, and pruritus.⁶ ACD is mostly caused by small molecules (<500 d), generally denoted as haptens. A variety of chemicals can act as haptens and cause ACD, including pharmaceutical,⁷ industrial (eg, hair dyes or fragrances),^{8,9} and metallic compounds (eg, nickel sulfate).¹⁰ These small molecules can easily penetrate the stratum corneum of the skin barrier and interact with cutaneous proteins to form covalent adducts. Most haptens are weak allergens, requiring repeated exposures before they cause sensitization. However, strong

allergens such urushiol (the allergenic component of poison ivy) can produce sensitization after a single exposure.¹¹

In addition to conventional ACD, there are also cases of systemic contact dermatitis, which is an inflammatory skin reaction occasionally seen as a flare up of previous dermatitis upon systemic exposure of allergen-sensitive individuals to the hapten (ie, administered orally, intravenously, or by inhalation). ¹² In one study, it was demonstrated that some subjects (13 out of 33) with a history of nickel-induced contact dermatitis develop dermatitis when challenged with nickel orally, while no subjects without a skin sensitivity to nickel exhibited a cutaneous reaction. ¹² These results suggest that cutaneous sensitization to a xenobiotic may result in a skin reaction upon subsequent systemic exposure to the same xenobiotic.

Delayed-type Hypersensitivity Reactions in the Skin After Systemic Drug Administration

DTH reactions in skin are also associated with the systemic administration of a wide number of drugs. Such drugs include antimicrobial (eg, sulfonamides, dapsone), anticonvulsant (eg, carbamazepine), anesthetic (eg, lidocaine), analgesic (eg, acetaminophen), antipsychotic (eg, clozapine), cardiovascular (eg, procainamide and hydralazine), nonsteroidal anti-inflammatory (eg, diclofenac), and steroid (eg, estrogen) agents. Naldi et al¹³ found that antimicrobial drugs are the most common agents associated with CDRs, followed by nonsteroidal anti-inflammatory drugs (NSAIDs) and other analgesics. However, CDRs in response to NSAID and analgesic therapy were more commonly associated with fatalities than those occurring with antimicrobial agents. These investigators also found that the most frequently reported serious antimicrobial induced-CDRs were angioedema, erythema multiforme, and Stevens Johnson Syndrome.

DTH reactions in skin observed after systemic drug administration generally occur 7 to 10 days after initiation of therapy and are commonly associated with fever. ¹⁴ While rash is the most common manifestation observed with these reactions, systemic organ involvement (eg, liver and kidney) is also observed. ¹⁵ Most CDRs rapidly subside upon removal of the offending agent. In some instances, however, drug rashes progress despite discontinuance of the drug.

Accurate causality assessment is an important issue for clinicians and scientists using human subjects to investigate the mechanisms of CDRs. 16,17 As many patients experiencing an acute skin eruption are receiving several drugs, identification of the causative agent is problematic. The ideal "proof" that an agent caused the skin eruption in a given patient is reappearance of the reaction upon re-exposure to the suspected agent. However, in most patients therapeutic need does not justify the risk of re-exposure. Hence, definitive proof of the offending agent is most commonly lacking. As a consequence, diagnosis is based on probability assess-

ment using the clinical history, timing of administration, and knowledge of previous associations of similar reactions with the suspected drug.¹⁷

EVIDENCE SUGGESTING THE INVOLVEMENT OF THE IMMUNE SYSTEM IN CUTANEOUS DRUG REACTIONS

Numerous lines of evidence, mainly derived from immunohistochemical analysis of skin lesions and generation of drug specific T-cell clones from individuals with a history of a CDR, indicate that the cell-mediated component of the immune system plays a central role in the pathogenesis of drug-induced DTH reactions in the skin. 18-20 Moreover, the delayed onset, swelling of lymph nodes, 21 and secretion of pro-inflammatory cytokines 22,23 during these reactions provide further support for the direct involvement of the immune system in these CDRs. Suppression of drug-induced hypersensitivity reactions by gradual dose escalation, which may be explained by immune tolerance, also suggests a direct involvement of the immune system in the pathogenesis of drug-induced DTH. 3

Drug-induced skin reactions appear most frequently as exanthemas (a widespread rash or eruption). Using skin-tests and immunohistochemical analysis, Britschgi et al²⁴ demonstrated a massive infiltration of neutrophils in pustules and T cells in the dermis and epidermis from patients with exanthematous pustulosis. Immunohistological analysis of skin at the time of bromisovalum-induced CDR also showed infiltration of CD4+ and CD8+ lymphocytes in dermal and epidermal infiltrates, respectively.25 Histological analysis of other drug-induced exanthemas have also showed mild to moderate mononuclear cell infiltration in the skin. 18 These mononuclear cells are composed of CD3+ T cells with a predominance of CD4+ T cells in the perivascular dermis, whereas an equal number of CD4+ and CD8+ T cells appear in the dermato-epidermal junction zone and epidermis of skin.²⁶ It is, however, important to note that lymphocyte infiltration in skin or enlargement of lymph nodes is also a consequence of a cutaneous infection that triggers an immune response.²⁷ Hence, it is possible that the immunohistological evidence for immune involvement cited above is mediated by a concurrent infection rather than a direct response to the drug itself. Of interest, it appears that viral infection may itself be a predisposing factor for the development of CDRs.⁴

The most compelling evidence supporting the involvement of the immune system in CDRs is provided through the demonstration of the proliferation of drug-specific T lymphocytes obtained from patients with a history of CDR. Pichler and colleagues have been able to isolate drug-specific CD4+ and CD8+ T-cell patients with a history of CDR to sulfonamides, cephalosporins, and anticonvulsants. ²⁸⁻³⁰ T-cell clones could not be isolated from patients

with a drug ingestion history but no history of CDR. Subsequently, similar observations have been made for additional drugs by other investigators.³¹ Further support has been provided by Roujeau and colleagues, who have demonstrated the presence of CD8+ cytotoxic drug-specific T cells in blister fluid of patients suffering from toxic epidermal necrolysis.^{32,33} In addition, Schnyder et al³⁴ were able to demonstrate T cell-mediated killing of interferon-γ treated keratinocytes isolated from a patient with a history of a CDR during treatment with sulfamethoxazole. Together, these studies provide substantial evidence for the role of T cell-mediated immunity in DTH skin reactions to drugs.

CRITICAL STEPS IN THE PROVOCATION OF IMMUNE REACTIONS TO XENOBIOTICS IN THE SKIN

Substantial advances in our understanding of immune responses in the skin have been derived from the study of contact sensitizing agents.³⁵ These studies reveal a complex sequential series of events that involve the engagement of multiple cell types and modulation by a variety of endogenous chemical mediators (Figure 1). Of importance, most small molecules are not believed to be direct immunogens or antigens but instead must first bind (generally covalently) to cellular or extracellular proteins prior to recognition by key immune molecules. These haptens are taken up by antigen presenting cells (APCs) in the skin either by pinocytosis or by receptor-mediated endocytosis. Following hapten uptake, APCs upregulate expression of specific surface molecules (eg. major histocompatability complex [MHC]). costimulatory molecules (eg, CD80), and cytokines.³⁶ The primary professional APCs in skin are Langerhans cells (LC),^{37,38} which are resident dendritic cells (DC) that serve a sentinel function. In response to antigenic challenge, LC migrate to the lymph node carrying the hapten and present it to responsive T cells. Epidermal keratinocytes (KC) also play a pivotal role in the initiation and propagation of

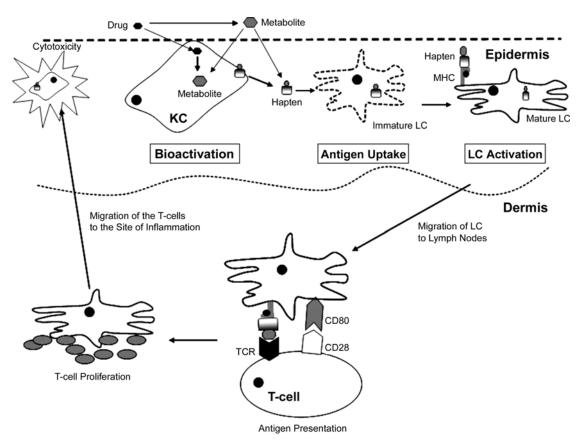


Figure 1. Schematic representation of critical events involved in xenobiotic-induced immune reactions in skin. Drug or metabolite enter the epidermis through the stratum corneum and diffuse into KC. Alternatively, drug or metabolite may enter via the systemic circulation. Drug may undergo bioactivation, followed by intracellular protein haptenation. Metabolite entering directly or released from KC may haptenate extracellular proteins. Antigen uptake by Langerhans cells,³⁷ together with inflammatory signals, results in activation/maturation of LC. Activation of LC results in the release of signals that direct migration of these cells to the draining lymph node. Upon arrival at the lymph node, LC present antigen to T cells in an MHC-restricted (major histocompatability) fashion. Engagement of the TCR in the presence of costimulatory signals (CD80/CD28) results in clonal expansion of xenobiotic-reactive T cells. These T cells express skin homing receptors, which play an important role in recruitment to the site of allergen presentation. Activation of xenobiotic-reactive T cells in the skin results in cell killing and inflammation.

immune responses in the skin.³⁹ Hence, an understanding of the mechanism of CDRs necessitates a consideration of critical events that may occur in the skin, which ultimately provoke the observed drug eruption.

Bioactivation

The preponderance of evidence suggests that formation of drug-protein adduct is the first critical step in the elicitation of an immune response following the administration of a small molecule. However, very few drug molecules are chemically reactive in nature, such that they are able to function as immunogens through direct binding to cellular proteins (eg, cephalosporin). Most drugs are chemically inert and must be metabolized (eg, to epoxides, quinones, or nitroso derivatives) or degraded (eg, penicillin) to reactive species prior to haptenation with cellular proteins. Indeed, a survey of drugs associated with DTH reactions in the skin reveals that all have been found to either be metabolized or degraded to reactive species (Table 1).

While the liver is quantitatively the primary site of bioactivation for most drugs, metabolism may also occur in other organs (eg, skin, kidney, or gut). Hence, drugs can distribute to the skin either as the parent compound or as preformed metabolite. Numerous drug metabolizing enzymes have been found to be present in the skin, including cytochromes P-450,⁵⁵ N-acetyltransferase,⁴⁸ flavin monooxygenases,⁵⁶ and cyclooxygenases.⁵⁷ We have demonstrated the ability of epidermal keratinocytes⁴⁸ and dermal fibroblasts (unpublished data, May 2005) to bioactivate sulfamethoxazole and/or dapsone, giving rise to intracellular haptenated

proteins. Evidence is currently lacking for the bioactivation in skin cells of other drugs associated with CDRs.

Dendritic Cell or Langerhans Cell Activation, Maturation, and Migration

Following entry into the skin, reactive drug or metabolite binds to cutaneous proteins and gives rise to drug-protein adducts, which may subsequently be processed and presented by DC/LC as neo-antigens (Figure 1). Activation of immature DC followed by subsequent migration to draining lymph nodes is a critical step for the induction of druginduced DTH reactions in the skin and other organs.⁵⁸ DC collect antigens in peripheral and visceral sites in the body, process these antigens, and transport them to the lymphoid organ in a well-synchronized fashion.^{59,60} During this journey to draining lymph nodes, DC undergo a maturation process.⁶¹ For the induction of DTH in skin, LC serve the primary role of APCs⁶² and are present both in the dermis and epidermis of the skin.63 Following interaction with an antigen (eg, contact allergen), LC become activated and migrate from skin to the regional draining lymph node. Various changes have been reported to occur in LC as a result of antigen exposure, including internalization of surface-MHC-II molecules via endocytosis, 64,65 induction of tyrosine phosphorylation, 66 modulation of various cell surface markers, 67,68 and expression of cytokines. 69,70

Processing and presentation of antigen by LC is a critical step in determining the nature of the immune response. Antigen processing involves the breakdown of long complex proteins into relatively shorter peptides to which the

Table 1. Drugs Known to Cause Cutaneous Drug Reactions That Have Also Been Shown to Form Chemically Reactive Metabolites	Table 1	l. Drugs Known to Cause	Cutaneous Drug Reactions	s That Have Also Been S	Shown to Form Chemica	llv Reactive Metabolites*
---	---------	-------------------------	--------------------------	-------------------------	-----------------------	---------------------------

Drug	Metabolite	Enzyme	Reference
Carbamazepine	Arene oxide Quinoneimine	CYP3A4	41,42
Sulfamethoxazole	Hydroxylamine Nitroso N-chloro	CYP2C9, CYP2E1, CYP3A4, myeloperoxidase	43-45
Dapsone	Hydroxylamine Nitroso N-chloro	CYP2C9, CYP2E1, CYP3A4, myeloperoxidase	45-48
Diclofenac	Acyl glucoronide	CYP2C9	49,50
Lidocaine	3-hydroxylidocaine	CYP2D6, CYP3A4	51
Abacavir	Aldehyde	Alcohol dehydrogenase	52
Phenytoin	Arene oxide Quinone	CYP2C9, CYP3A4	53
Procainamide	Hydroxylamine Nitroso	Unknown	54

^{*}Adapted and modified from Park et al.40

hapten may be attached.⁷¹⁻⁷³ This breakdown enables the peptide to bind loosely (noncovalently) to the MHC within the cell. Antigen disposition is the key step that determines which MHC molecule will present the antigens. Extracellular antigens (eg, contact allergens), which involve capture of antigen in the endosome followed by lysosomal degradation, are expected to be presented via class II MHC. Intracellular antigens (eg, drug-protein adducts formed intracellularly) appear to be preferentially presented in a class I MHC-restricted manner.⁷⁴ Herouet et al⁷⁵ showed a specific increase of MHC-II expression in LC following application of the contact sensitizer dinitrobenzene sulfonic acid in mice. Hence, upregulation of these antigen-presenting molecules upon exposure to allergens appears to play an important role in mediating DTH reactions.

The critical role played by LC in the development of an immune response requires highly synchronized changes in both phenotype and functionality, including cell-cell interactions. These changes have been shown to be regulated largely by cytokines and other pro-inflammatory signals, such nitric oxide (NO) or reactive oxygen species (ROS). Recently, evidence has been presented that indicates that alterations in DC redox balance induced by chemical allergens may play an important role in triggering the maturation of LC. The potential role of such mediators in CDRs is discussed further in a subsequent section of this review.

Expression of Costimulatory Molecules

After interaction with antigen, LC migrate to the lymph node and present the antigen-MHC complex to responsive T cells (Figure 1). T-cell activation requires at least 2 signals. The first signal is delivered in the form of T-cell receptor (TCR) engagement with peptide/MHC expressed on the surface of an APC. The second signal is provided by costimulatory molecules on the APC interacting with their respective receptors on the engaged T cell. The most widely studied costimulatory molecules B7-1 (CD80) and B7-2 (CD86) bind to their respective receptors on T cells (CD28 and CTLA-4 or CD152), which determines the activation state of the T cell.^{78,79} In addition to these well-established costimulatory molecules, recent work has identified additional costimulatory molecules (eg, CD40-CD154, OX-40-OX-40L, RANK-RANKL, and PD-1-PD-L-1).80 Of importance, LC have been found to express various costimulatory molecules, including CD80 and CD86.81 In addition, KC are also known to constitutively express several costimulatory molecules (eg, CD80), which are upregulated in response to contact-allergen exposure.82

Presentation of peptide-MHC complex by DC/LC to T cells can either lead to activation or tolerance, depending upon the expression of costimulatory molecules. Antigen presentation by steady-state DC/LC lacking full expression of

costimulatory molecules fail to drive T cells toward productive activation. Important insight into the significance of costimulatory molecule expression during antigen presentation was recently provided by Hochweller and Anderton, 80 who examined the kinetics of costimulatory molecule expression by both DC and T cells in a study of tolerance versus activation. These investigators demonstrated that CD154, OX-40, PD-1, and RANK-L were all expressed on T cells after administration of either soluble antigen (peptide-ovalbumin) or soluble antigen in presence of lipopolysaccharide (LPS). Higher and longer expression of CD154 and OX-40 were observed during the induction of immunity compared with tolerance. However, increases in CD40, RANK, PDL-1, CD80, and CD86 were only observed after LPS in addition to antigen administration. These data indicate that until and unless sufficient expression of costimulatory molecules on DC occurs, T cells are not activated to generate an immune response.

T-Cell Propagation

As described above, LC present antigens to T cells in the lymphatic organs in an MHC-restricted fashion. Antigen encounter and recognition activates the transition of a naive T cell to a memory/effector T cell.⁸³ As illustrated in Figure 1, clonal expansion of drug-specific T cells gives rise to a population of T cells with skin homing receptors that localize to the skin where effector function is needed.

Detection of drug-specific T cells in peripheral blood and inflamed tissues^{84,85} from patients with drug hypersensitivity gives evidence of a direct involvement of T cells in DTH reactions associated with drugs. Several investigators have shown that various drugs and or their metabolites (eg, carbamazepine, sulfamethoxazole, lamotrigine, lidocaine, and mepivacaine) can stimulate T cells via interaction with the TCR in an MHC-restricted manner.^{31,85-88} The interaction of these drugs with TCR causes cytokine secretion, proliferation, or cytotoxicity in the reactive T-cell clones.

Recently, Gerber and Pichler have provided evidence for T-cell activation arising from noncovalent drug-T cell interactions.⁸⁹ This finding has led to a reconsideration of the essential role of haptenation by small molecules and is discussed in depth in a companion article in this series.⁹⁰

The means by which drug-specific T cells are recruited to the site of antigen entry is an area of ongoing investigation. It has been demonstrated that T cells infiltrating the skin express a unique skin homing receptor known as cutaneous lymphocyte-associated antigen (CLA).^{91,92} CLA is a carbohydrate epitope present in skin homing T cells, which binds specifically to E-selectin. CLA+ T cells isolated from nickel-sensitive patients have been shown to be activated in response to nickel exposure, whereas CLA- T cells were not activated.⁹¹ Binding of CLA and E-selectin, which is

important for trafficking of T cells through the activated endothelium, also involves the interaction of additional surface markers.⁹²

Most drug-induced DTH reactions exhibit complex and overlapping cytokine/chemokine profiles, where various T cells with distinct functions contribute to the clinical manifestations. Immunohistological data suggest that the pathology of drug-induced skin eruptions may depend on the grade of CD4+ and/or CD8+ T-cell activation and function. Is In maculopapular or eczematous drug eruptions, the immune response appears to be mediated via CD4+ T-cell activation and MHC-II -restricted drug presentation, Is whereas CD8+ T-cell activation results in more severe skin symptoms (bullous skin disease). These T cells are associated with distinct cytokine secretion profiles (discussed below).

POTENTIAL ROLE OF KERATINOCYTES IN ANTIGEN PRESENTATION

KC play an important role in the initiation and propagation of immune responses in the skin. While they do not possess the ability to present antigen under basal conditions, KC may do so in the presence of inflammatory stimuli. He ability to present antigen may play an important role in the targeting of KC for killing by cytotoxic T cells recruited to the skin in CDRs. Presentation of antigens by KC to T cells necessitates the ability of KC to perform numerous APC-like functions, including antigen uptake, intracellular processing, and presentation in the context of MHC molecules, as well as providing reciprocal interaction of cell surface receptors for costimulatory molecules between T cells and KC.

Studies have demonstrated that KC are able to express both types of major histocompatability complexes (MHC-I and MHC-II). 95,96 While MHC-I is expressed constitutively on the surface of KC, MHC-II is only expressed in response to a pro-inflammatory insult (eg. IFN-y exposure). 95 It has also been demonstrated that contact allergens induce the expression of the important costimulatory molecule, CD80, in KC.82 Moreover, expression of intercellular adhesion molecule (ICAM-1) is upregulated in the presence of known contact allergens. 97,98 ICAM-1 is known to mediate antigen presentation in functional APC via interaction with the lymphocyte function-associated antigens (LFA-1)99 and may exhibit similar effects in KC. While further investigation into MHC-restricted antigen presentation by KC is needed, evidence to date suggests that antigen presentation by these cells may play an important role in mediating CDRs.

POTENTIAL MEDIATORS OF DELAYED-TYPE HYPERSENSITIVITY REACTIONS IN THE SKIN

Cytokine/Chemokines

The cutaneous immune response to drugs is regulated in large part by cytokines/chemokines. Cutaneous cells secrete

a wide variety of cytokines/chemokines, many of which are induced by allergen exposure and are pro-inflammatory in nature (tumor necrosis factor α [TNF- α], interleukin [IL]-1, IL-2, and IL-6). 100-102 In addition, neutrophil attractant (IL-8), growth promoting (IL-6, Il-7, IL-15, and granulocyte colony stimulating factor [GM-CSF]), and anti-inflammatory cytokines (IL-4, IL-10) are also secreted in the cutaneous environment. 103,104 Drug specific T cells are also believed to mediate inflammatory skin reactions through the release of various chemokines (eg, RANTES, eotaxin, or IL-8).93 Several of these cytokines (ie, TNF-α, IL-1β, and IL-18) have been found to be important for the mobilization and migration of LC following exposure to contact allergens. $^{76,105-107}$ In addition, IFN- γ and TNF- α have been found in blister fluid from patients of toxic epidermal necrolysis. 108 The observation that these cytokines were present in much higher concentrations in blister fluids than infiltrating mononuclear cells suggests that they are secreted by skin cells (eg, KC) rather than cells recruited to the site. 108 The exact role of cytokines in regulating cell killing, however, remains to be determined.

As described previously, pro-inflammatory cytokines (eg, TNF- α) are among the important signals that upregulate the expression of costimulatory molecules on the surface of APC. ¹⁰⁹ Pro-inflammatory cytokines also result in the upregulation of MHC-II in KC, which enables these cells to present antigen in an MHC-II-dependent fashion. ^{110,111} Numerous contact allergens (eg, nickel, dinitrofluorobenzene) have been reported to induce cytokines (eg, TNF- α , IL-1) in cultured KC. ^{112,113} Pro-inflammatory cytokines are also induced in epidermal KC and dermal infiltrates from mice following exposure to the allergen trinitrochlorobenzene. ¹¹⁴ Cytokines secreted by KC in vitro are used by some investigators to evaluate the sensitizing potential of small molecular weight compounds. ¹¹⁵

Evidence to date indicates that the functional characteristics of drug-specific T cells is determined by distinct cytokine profiles. Two major phenotypes of cytokine secreting T cells have been identified: type-1, which secrete IFN-γ, and type-2, which secrete IL-4, IL-5, IL-10, and IL-13. To Generally, type-1 T cells are involved in pro-inflammatory effector activities, while type-2 induce allergic responses. ACD is predominantly associated with type-1 T cells with less but significant involvement of type-2 cells. The balance between these 2 types of T cells depends on various factors, including the concentration of the antigen, type of antigen, and the cytokines involved in the early interaction of the T cell and the DC. 120-122

Potent contact allergens are found to activate IFN- γ secreting type-1 effector T cells, and it appears that IFN- γ plays a major role in propagating the cutaneous inflammatory response. Moreover, neutralization of IFN- γ (using monoclonal antibody) at the time of the challenge with the allergen

(eg, picryl chloride) strongly reduces inflammation. ¹²³ In patients with drug-induced DTH, drug-activated T cells secreted high amounts of type-2 cytokines with normal or low levels of type-1 cytokines. ²⁸

Nitric Oxide

Nitric oxide (NO) is a small regulatory molecule known to play diverse roles in a wide variety of biological phenomena that can be protective, regulatory, or toxic to cells depending on its intracellular concentration. ¹²⁴ NO has been implicated in the pathogenesis of various immune-mediated skin diseases. NO is known to exert its effect at multiple cellular and molecular levels that include direct effects on different immune cell types (eg, DC, T cells, neutrophils, or mast cells) either at the site of inflammation or in the lymph node. ¹²⁵ NO has also been reported to regulate expression of different cytokines and cell surface markers that are essential for the cell-mediated immune response. ¹²⁶ While the role of NO in cutaneous physiology has been recently reviewed, ¹²⁷ there has been little consideration of its potential role in mediating CDRs.

NO is synthesized by 3 different enzymes commonly known as inducible NO-synthase (iNOS), ¹²⁸ endothelial NOS (eNOS), or neuronal NOS. Among these 3 enzymes, iNOS is known to be responsible for the high output NO production in different cell types following inflammatory insults. ¹²⁹ Of importance, studies have demonstrated that iNOS is expressed in various types of skin cells, including KC, ¹³⁰ LC, ¹³¹ melanocytes, ¹³² and dermal fibroblasts. ¹²⁹ KC have also been reported to contain constitutive NOS, which can be activated upon exposure to various stimuli (eg, ultraviolet exposure). ^{128,133} Moreover, the presence of eNOS has also been demonstrated in the dermal vasculature of skin. ¹³⁴

Skin biopsies of patients with actively progressing Stevens Johnson Syndrome (SJS) and toxic epidermal necrolysis (TEN) has shown an upregulation of iNOS in KC and infiltrating inflammatory cells as demonstrated by immunohistochemistry and reverse transcription polymerase chain reaction.¹³⁵ It has been suggested that the high amount of NO produced by upregulated iNOS may play a role in the massive epidermal cell death observed in patients with SJS and TEN. In addition to a potential role in epidermal apoptosis and/or necrosis in SJS and TEN, NO is believed to play a regulatory role in elicitation of a cutaneous immune response. For example, immunohistological analysis of skin biopsies from patients with ACD demonstrated expression of iNOS in the upper dermal microvasculature of involved sites. 136 These studies suggest a potential role for iNOS and its product NO in DTH reactions in the skin.

In agreement with these clinical findings, other investigators 137,138 found that the NOS inhibitor L-nitroarginine methyl ester (L-NAME) suppressed allergen-induced con-

tact hypersensitivity (CHS) in mice. Other investigators, however, have not supported these findings. ¹⁰⁵ Moreover, the CHS response to dinitrofluorobenzene is enhanced in iNOS-deficient mice compared with healthy controls, suggesting iNOS may normally downregulate the inflammatory response. ¹²⁵ Recent reports have shown that in vivo encounter with contact allergens upregulates iNOS expression in both KC and LC. ¹³⁹ However, it is not clear whether this activation of iNOS is a direct consequence of allergen contact or a downstream event. As described previously, exposure to contact allergens causes secretion of cytokines from KC and LC, which are known to upregulate iNOS expression. ^{140,141} Hence, the observed iNOS induction may be secondary to the release of pro-inflammatory cytokines.

NO, in combination with cytokines secreted from KC/LC following contact allergen exposure, has been suggested to direct various effector cells (eg, T cells, macrophages, neutrophils) to the site of inflammation. 125 Of interest, it has also been demonstrated that intracellular concentration of NO could be a crucial factor in determining its role to direct neutrophils. Wanikiat et al 142 have shown that low NO concentration facilitates chemotaxis of human neutrophils, while high concentration of NO inhibited chemotaxis. Of importance, NO is also known to downregulate expression of several surface molecules (eg, ICAM-1, vascular adhesion molecule [VCAM-1]) that are known to mediate adhesion of leukocytes to vascular endothelial cells and infiltration of leukocytes into the peripheral tissues. 143

Interesting insight into the potential impact of cutaneous NO production has been provided through the demonstration that NO induces apoptosis of DC and T cells. 144,145 Apoptosis of antigen-presenting DC in the skin or draining lymph nodes would be expected to limit the expansion of antigen-specific T cells, 144 thus downregulating the immune response. NO produced by DC and macrophages in the skin would also potentially induce apoptosis in recruited drugspecific T cells. 146

Thus, while biopsy samples from patients with acute CDR suggest a potential role for NO/NOS in modulating these reactions, the results of experiments in animal models of CHS are conflicting. While further experimental work is needed to ascertain what role, if any, this potential mediator plays in the mechanism of CDRs, the cumulative evidence to date suggests a role for NO/iNOS in downregulating the cutaneous inflammatory response to antigens.

Reactive Oxygen Species

Reactive oxygen species (ROS) are known to be engaged in several of the critical steps (eg, DC activation/maturation, release of cytokines, T-cell activation) involved in the skin immune response.¹²⁰ At sites of inflammation, tissue damage has been linked to the release of various pro-oxidants, ¹⁴⁷

which have the potential to alter DC phenotype/function. These pro-oxidants may either directly affect DC maturation and/or activation¹⁴⁸ or act as a signal to induce expression of different immune regulatory molecules (eg, ICAM-1,¹⁴⁹ MHC-II,¹⁵⁰ cytokines¹⁵¹). Moreover, altered antioxidant status in mononuclear blood cells from patients with CDRs¹⁵² suggests a direct involvement of ROS during DTH reactions. Thiobarbituric acid derivatives and carbonyl content (indicators of ROS-mediated cellular oxidative stress) were also found to be elevated in the serum of those patients with CDRs to phenytoin and carbamazepine.¹⁵²

Human immunodeficiency virus (HIV)-infected individuals have been found to be more susceptible to drug-induced cutaneous DTH reactions. 153 A lowered antioxidant (eg. glutathione) status in HIV-infected individuals has been suggested as a predisposing factor in the higher frequency of CDR in this population.¹⁵⁴ Altered antioxidant levels (measured as higher oxidized glutathione) in skin biopsies of ACD patients^{155,156} and irreversible inhibition of mammalian thioredoxin reductase by the contact allergen dinitrochlorobenzene¹⁵⁷ also suggest involvement of ROS in CDRs. Exposure to reactive metabolites of sulfamethoxazole and dapsone, drugs known to cause CDRs, have been shown to cause depletion of reduced glutathione, 48 as well as higher ROS generation in epidermal KC158 and dermal fibroblasts (unpublished data, May 2005). Taken together, these data indicate that reduced antioxidant status and/or higher ROS generation in skin cells may predispose individuals to CDRs.

Among various ROS, superoxide has been shown to induce early maturation of (monocyte-derived) DC via upregulation of cell surface markers (CD80, CD83, and CD86) and to downregulate endocytic activity. Superoxide-induced DC maturation is inhibited in the presence of the antioxidant N-acetylcysteine. In contrast, H₂O₂ failed to cause DC maturation and CD86 upregulation. H₂O₂ has been found to upregulate cytokines (eg, TNF-α, IL-8) by human DC, suggesting that these cells could convey the immune response through enhanced cytokine production as a consequence of oxidative stress.

ROS have also been shown to increase the expression of numerous adhesion molecules (eg, E-selectin, ICAM-1, and VCAM-1). Of importance, N-acetylcysteine reduces ROS-induced ICAM-1 expression in human KC. Expression of E-selectin and VCAM-1 by contact allergens is also prevented in the presence of NAC. These results suggest that ROS are able to regulate the recruitment of T cells to the site of inflammation via alterations in the expression of adhesion molecules.

ROS is also known to influence the DC-T cell cross-talk, one of the most important criteria for antigen presentation. Interaction between DC and T cells is a bidirectional

phenomenon, in which DC deliver activation signals to T cells and also receive signals back from the responding T cells, thereby undergoing terminal maturation. Interference in the cellular redox regulation pathways by ROS has been found to influence this bidirectional communication between DC and T cells. ¹⁶³ In particular, both DC and T cells exhibited an elevated level of ROS following antigen-specific interactions, which was blocked in the presence of an antioxidant (ebselen). Moreover, ebselen also inhibited the DC-induced proliferation and cytokine production by T cells. These results imply that endogenously produced ROS in DC and T cells act as second messengers, which play an important role in altering their cellular functions during antigen presentation.

ROS has been demonstrated to induce cell death in T cells, which can be prevented by the application of antioxidants. ¹⁶⁴ Higher levels of ROS are known to cause necrosis, whereas lower ROS levels induce apoptosis. ¹⁶⁵ Apoptosis of T lymphocytes triggered by ROS has been shown to be mediated via different cellular processes, including lysosomal membrane destabilization, ¹⁶⁶ Fas-ligand activation, ¹⁶⁷ and downregulation of Bcl-2. ¹⁶⁸

Although numerous studies have suggested a direct involvement of ROS in the elicitation of immune responses in various DTH reactions, studies have failed to show significant inhibition of ACD by topical or systemic application of antioxidants. 169,170 This discrepancy can be explained by the fact that different ROS may simultaneously trigger different cellular processes in various skin cell types. For example, elevated ROS may cause MHC-II upregulation in DC, 150 but at the same time it may induce apoptosis of T cells 166 and thus provide antagonizing effects. Future research should be directed at elucidating the cell-specific mechanism of ROS generation and function during druginduced immune reactions in skin.

CONCLUSION

Studies examining the immune response to compounds applied epicutaneously have provided important insight into the mechanism of immune responses to xenobiotics in the skin. These studies have revealed several critical steps that must take place prior to the development of a measurable inflammatory response (ie, a rash). But what do these studies tell us about the mechanism by which drugs administered systemically provoke reactions in the skin? It is clear that the pattern of rash observed in response to systemically administered drugs is usually quite different from that provoked by contact sensitizing agents. However, numerous lines of evidence indicate that the immunological responses display significant similarity. As described in this review, xenobiotic-specific reactive T cells have been isolated from patients with ACD and various forms of CDRs. In addition,

upregulation of important surface molecules known to mediate lymphocyte trafficking and activation have been observed in the skin after epicutaneous administration of contact sensitizers and in biopsies of lesions from patients with CDRs. The congruence of these immunological responses suggests important similarities in the mechanism of ACD and CDRs, providing a rationale for the use of ACD as an important experimental paradigm for mechanistic insight that may then be tested within the experimental and ethical limitations surrounding studies in patients with CDRs. It is anticipated that elucidation of the key mediators of these reactions will provide therapeutic targets for the prevention and management of CDRs. Moreover, the available mechanistic insight should foster the development of preclinical screening approaches to identify those compounds with a high likelihood of provoking such reactions.

ACKNOWLEDGMENTS

This study was supported in part by grant GM63821 from the National Institutes of Health (NIH), Bethesda, MD.

REFERENCES

- 1. Pirmohamed M, Breckenridge A, Kitteringham N, et al. Adverse drug reactions. *BMJ*. 1998;316:1295-1298.
- 2. Vervloet D, Durham S. Adverse reactions to drugs. *BMJ*. 1998;316:1511-1514.
- 3. Naisbitt DJ. Drug hypersensitivity reactions in skin: understanding mechanisms and the development of diagnostic and predictive tests. *Toxicology*. 2004;194:179-196.
- 4. Svensson CK, Cowen EW, Gaspari AA. Cutaneous drug reactions. *Pharmacol Rev.* 2001;53:357-379.
- 5. Romano A, Torres MJ, Quaratino D, et al. Diagnostic evaluation of delayed hypersensitivity to systematically administered drugs. *Allergy*. 1999;54:23-27.
- 6. Saint-Mezard P, Rosieres A, Krasteva M, et al. Allergic contact dermatitis. *Eur J Dermatol*. 2004;14:284-295.
- 7. Sertoli A, Francalanci S, Acciai MC, et al. Epidemiological survey of contact dermatitis in Italy (1984-1993) by GIRDCA (Gruppo Italiano Ricerca Dermatiti da Contatto e Ambientali). *Am J Contact Dermat.* 1999;10:18-30.
- 8. Matulich J, Sullivan J. A temporary henna tattoo causing hair and clothing dye allergy. *Contact Dermatitis*. 2005;53:33-36.
- 9. Militello G, James W. Lyral: a fragrance allergen. Dermatitis. 2005;16:41-44.
- 10. Bonamonte D, Foti C, Antelmi AR, et al. Nickel contact allergy and menstrual cycle. *Contact Dermatitis*. 2005;52:309-313.
- 11. Li LY Jr, Cruz PD Jr. Allergic contact dermatitis: pathophysiology applied to future therapy. *Dermatol Ther*. 2004;17:219-223.
- 12. Jensen C, Lisby S, Larsen J, et al. Characterization of lymphocyte subpopulations and cytokine profiles in peripheral blood of nickelsensitive individuals with systemic contact dermatitis after oral nickel exposure. *Contact Dermatitis*. 2004;50:31-38.
- 13. Naldi L, Conforti A, Venegoni M, et al. Cutaneous reactions to drugs: an analysis of spontaneous reports in 4 Italian regions. *Br J Clin Pharmacol*. 1999;48:839-846.

- 14. Shepherd GM. Hypersensitivity reactions to drugs: evaluation and management. *Mt Sinai J Med*. 2003;70:113-125.
- 15. Knowles SR, Shapiro LE, Shear NH. Anticonvulsant hypersensitivity syndrome: incidence, prevention, and management. *Drug Saf.* 1999;21:489-501.
- 16. Aronson JK, Ferner RE. Joining the DoTS: new approach to classifying adverse drug reactions. *BMJ*. 2003;327:1222-1225.
- 17. Edwards IR, Aronson JK. Adverse drug reactions: definitions, diagnosis, and management. *Lancet*. 2000;356:1255-1259.
- 18. Hari Y, Frutig-Schnyder K, Hurni M, et al. T cell involvement in cutaneous drug eruptions. *Clin Exp Allergy*. 2001;31:1398-1408.
- 19. Hertl M, Merk HF. Lymphocyte activation in cutaneous drug reactions. *J Invest Dermatol*. 1995;105:95S-98S.
- 20. Pichler WJ, Schnyder B, Zanni MP, et al. Role of T cells in drug allergies. *Allergy*. 1998;53:225-232.
- 21. Bessmertny O, Hatton R, Gonzalez-Peralta R. Antiepileptic hypersensitivity syndrome in children. *Ann Pharmacother*. 2001;35:533-538.
- 22. Yawalkar N. Drug-induced exanthems. *Toxicology*. 2005;209:131-134.
- 23. Merk HF. Diagnosis of drug hypersensitivity: lymphocyte transformation test and cytokines. *Toxicology*. 2005;209:217-220.
- 24. Britschgi M, Steiner UC, Schmid S, et al. T-cell involvement in drug-induced acute generalized exanthematous pustulosis. *J Clin Invest*. 2001;107:1433-1441.
- 25. Miyauchi H, Hosokawa H, Akaeda T, et al. T-cell subsets in drug-induced toxic epidermal necrolysis: possible pathogenic mechanism induced by CD8-positive T cells. *Arch Dermatol*. 1991;127:851-855.
- 26. Pichler WJ. T cells in drug allergy. *Curr Allergy Asthma Rep.* 2002;2:9-15.
- 27. Kaplan MH, Hall WW, Susin M, et al. Syndrome of severe skin disease, eosinophilia, and dermatopathic lymphadenopathy in patients with HTLV-II complicating human immunodeficiency virus infection. *Am J Med.* 1991;91:300-309.
- 28. Mauri-Hellweg D, Bettens F, Mauri D, et al. Activation of drug-specific CD4+ and CD8+ T cells in individuals allergic to sulfonamides, phenytoin, and carbamazepine. *J Immunol*. 1995;155:462-472.
- 29. Schnyder B, Burkhart C, Schnyder-Frutig K, et al. Recognition of sulfamethoxazole and its reactive metabolites by drug-specific CD4+ T cells from allergic individuals. *J Immunol*. 2000;164:6647-6654.
- 30. Zanni MP, von Greyerz S, Schnyder B, et al. HLA-restricted, processing- and metabolism-independent pathway of drug recognition by human alpha beta T lymphocytes. *J Clin Invest*. 1998;102:1591-1598.
- 31. Naisbitt DJ, Farrell J, Wong G, et al. Characterization of drug-specific T cells in lamotrigine hypersensitivity. *J Allergy Clin Immunol*. 2003;111:1393-1403.
- 32. Nassif A, Bensussan A, Boumsell L, et al. Toxic epidermal necrolysis: effector cells are drug-specific cytotoxic T cells. *J Allergy Clin Immunol.* 2004;114:1209-1215.
- 33. Nassif A, Bensussan A, Dorothee G, et al. Drug specific cytotoxic T cells in the skin lesions of a patient with toxic epidermal necrolysis. *J Invest Dermatol.* 2002;118:728-733.
- 34. Schnyder B, Frutig K, Mauri-Hellweg D, et al. T-cell-mediated cytotoxicity against keratinocytes in sulfamethoxazol-induced skin reaction. *Clin Exp Allergy*. 1998;28:1412-1417.
- 35. Kimbe I, Basketter DA, Gerberick GF, et al. Allergic contact dermatitis. *Int Immunopharmacol*. 2002;2:201-211.

- 36. De Smedt AC, Van Den Heuvel RL, Van Tendeloo VF, et al. Capacity of CD34+ progenitor-derived dendritic cells to distinguish between sensitizers and irritants. *Toxicol Lett.* 2005;156:377-389.
- 37. Pichler WJ, Tilch J. The lymphocyte transformation test in the diagnosis of drug hypersensitivity. *Allergy*. 2004;59:809-820.
- 38. Romani N, Holzmann S, Tripp CH, et al. Langerhans cells -dendritic cells of the epidermis. *APMIS*. 2003;111:725-740.
- 39. Banerjee G, Damodaran A, Devi N, et al. Role of keratinocytes in antigen presentation and polarization of human T lymphocytes. *Scand J Immunol*. 2004;59:385-394.
- 40. Park BK, Kitteringham NR, Powell H, et al. Advances in molecular toxicology: towards understanding idiosyncratic drug toxicity. *Toxicology*. 2000;153:39-60.
- 41. Ju C, Uetrecht JP. Detection of 2-hydroxyiminostilbene in the urine of patients taking carbamazepine and its oxidation to a reactive iminoquinone intermediate. *J Pharmacol Exp Ther.* 1999;288:51-56.
- 42. Madden S, Maggs JL, Park BK. Bioactivation of carbamazepine in the rat in vivo: evidence for the formation of reactive arene oxide(s). *Drug Metab Dispos*. 1996;24:469-479.
- 43. Cribb AE, Miller M, Tesoro A, et al. Peroxidase-dependent oxidation of sulfonamides by monocytes and neutrophils from humans and dogs. *Mol Pharmacol*. 1990;38:744-751.
- 44. Cribb AE, Spielberg SP, Griffin GP. N4-hydroxylation of sulfamethoxazole by cytochrome P450 of the cytochrome P4502C subfamily and reduction of sulfamethoxazole hydroxylamine in human and rat hepatic microsomes. *Drug Metab Dispos*. 1995;23:406-414.
- 45. Uetrecht JP, Shear NH, Zahid N. N-chlorination of sulfamethoxazole and dapsone by the myeloperoxidase system. *Drug Metab Dispos*. 1993;21:830-834.
- 46. Winter HR, Wang Y, Unadkat JD. CYP2C8/9 mediate dapsone N-hydroxylation at clinical concentrations of dapsone. *Drug Metab Dispos*. 2000;28:865-868.
- 47. Mitra AK, Thummel KE, Kalhorn TF, et al. Metabolism of dapsone to its hydroxylamine by CYP2E1 in vitro and in vivo. *Clin Pharmacol Ther.* 1995;58:556-566.
- 48. Reilly TP, Lash LH, Doll MA, et al. A role for bioactivation and covalent binding within epidermal keratinocytes in sulfonamide-induced cutaneous drug reactions. *J Invest Dermatol*. 2000;114:1164-1173.
- 49. Yan Z, Li J, Huebert N, et al. Detection of a novel reactive metabolite of diclofenac: evidence for CYP2C9-mediated bioactivation via arene oxides. *Drug Metab Dispos*. 2005;33:706-713.
- 50. Kumar S, Samuel K, Subramanian R, et al. Extrapolation of diclofenac clearance from in vitro microsomal metabolism data: role of acyl glucuronidation and sequential oxidative metabolism of the acyl glucuronide. *J Pharmacol Exp Ther*. 2002; 303:969-978.
- 51. Masubuchi Y, Umeda S, Igarashi S, et al. Participation of the CYP2D subfamily in lidocaine 3-hydroxylation and formation of a reactive metabolite covalently bound to liver microsomal protein in rats. *Biochem Pharmacol*. 1993;46:1867-1869.
- 52. Walsh JS, Reese MJ, Thurmond LM. The metabolic activation of abacavir by human liver cytosol and expressed human alcohol dehydrogenase isozymes. *Chem Biol Interact*. 2002;142:135-154.
- 53. Cuttle L, Munns AJ, Hogg NA, et al. Phenytoin metabolism by human cytochrome P450: involvement of P450 3A and 2C forms in secondary metabolism and drug-protein adduct formation. *Drug Metab Dispos*. 2000;28:945-950.

- 54. Uetrecht JP. Reactivity and possible significance of hydroxylamine and nitroso metabolites of procainamide. *J Pharmacol Exp Ther*. 1985;232:420-425.
- 55. Swanson HI. Cytochrome P450 expression in human keratinocytes: an aryl hydrocarbon receptor perspective. *Chem Biol Interact*. 2004;149:69-79.
- 56. Janmohamed A, Dolphin CT, Phillips IR, et al. Quantification and cellular localization of expression in human skin of genes encoding flavin-containing monooxygenases and cytochromes P450. *Biochem Pharmacol*. 2001;62:777-786.
- 57. Lee JL, Mukhtar H, Bickers DR, et al. Cyclooxygenases in the skin: pharmacological and toxicological implications. *Toxicol Appl Pharmacol*. 2003;192:294-306.
- 58. Banchereau J, Steinman RM. Dendritic cells and the control of immunity. *Nature*. 1998;392:245-252.
- 59. Randolph GJ. Dendritic cell migration to lymph nodes: cytokines, chemokines, and lipid mediators. *Semin Immunol*. 2001;13:267-274.
- 60. Randolph GJ, Angeli V, Swartz MA. Dendritic-cell trafficking to lymph nodes through lymphatic vessels. *Nat Rev Immunol*. 2005;5:617-628.
- 61. Kimber I, Cumberbatch M. Dendritic cells and cutaneous immune responses to chemical allergens. *Toxicol Appl Pharmacol*. 1992;117:137-146.
- 62. Kimber I, Cumberbatch M, Betts CJ, et al. Dendritic cells and skin sensitization hazard assessment. *Toxicol In Vitro*. 2004;18:195-202.
- 63. Shortman K, Liu YJ. Mouse and human dendritic cell subtypes. *Nat Rev Immunol.* 2002;2:151-161.
- 64. Becker D, Mohamadzadeh M, Reske K, et al. Increased level of intracellular MHC class II molecules in murine Langerhans cells following in vivo and in vitro administration of contact allergens. *J Invest Dermatol*. 1992;99:545-549.
- 65. Girolomoni G, Simon JC, Bergstresser PR, et al. Freshly isolated spleen dendritic cells and epidermal Langerhans cells undergo similar phenotypic and functional changes during short-term culture. *J Immunol.* 1990;145:2820-2826.
- 66. Neisius U, Brand P, Plochmann S, et al. Detection of increased tyrosine phosphorylation in murine Langerhans cells after stimulation with contact sensitizers. *Arch Dermatol Res.* 1999;291:22-27.
- 67. Aiba S, Katz SI. Phenotypic and functional characteristics of in vivo-activated Langerhans cells. *J Immunol*. 1990;145:2791-2796.
- 68. Verrier AC, Schmitt D, Staquet MJ. Fragrance and contact allergens in vitro modulate the HLA-DR and E-cadherin expression on human epidermal Langerhans cells. *Int Arch Allergy Immunol*. 1999;120:56-62.
- 69. Enk AH, Katz SI. Early molecular events in the induction phase of contact sensitivity. *Proc Natl Acad Sci USA*. 1992;89:1398-1402.
- 70. Wang B, Feliciani C, Howell BG, et al. Contribution of Langerhans cell-derived IL-18 to contact hypersensitivity. *J Immunol*. 2002;168:3303-3308.
- 71. Kessler BM, Glas R, Ploegh HL. MHC class I antigen processing regulated by cytosolic proteolysis-short cuts that alter peptide generation. *Mol Immunol*. 2002;39:171-179.
- 72. Cresswell P. Assembly, transport, and function of MHC class II molecules. *Annu Rev Immunol*. 1994;12:259-293.
- 73. Cresswell P, Androlewicz MJ, Ortmann B. Assembly and transport of class I MHC-peptide complexes. *Ciba Found Symp*. 1994;187:150-162.

- 74. Park BK, Pirmohamed M, Kitteringham NR. Role of drug disposition in drug hypersensitivity: a chemical, molecular, and clinical perspective. *Chem Res Toxicol*. 1998;11:969-988.
- 75. Herouet C, Cottin M, LeClaire J, et al. Contact sensitizers specifically increase MHC class II expression on murine immature dendritic cells. *In Vitr Mol Toxicol*. 2000;13:113-123.
- 76. Cumberbatch M, Dearman RJ, Griffiths CE, et al. Epidermal Langerhans cell migration and sensitization to chemical allergens. *APMIS*. 2003;111:797-804.
- 77. Mizuashi M, Ohtani T, Nakagawa S, et al. Redox imbalance induced by contact sensitizers triggers the maturation of dendritic cells. *J Invest Dermatol.* 2005;124:579-586.
- 78. Chambers CA, Allison JP. Costimulation in T cell responses. *Curr Opin Immunol*. 1997;9:396-404.
- 79. McAdam AJ, Schweitzer AN, Sharpe AH. The role of B7 costimulation in activation and differentiation of CD4+ and CD8+ T cells. *Immunol Rev.* 1998;165:231-247.
- 80. Hochweller K, Anderton SM. Kinetics of costimulatory molecule expression by T cells and dendritic cells during the induction of tolerance versus immunity in vivo. *Eur J Immunol*. 2005;35:1086-1096.
- 81. Yokozeki H, Takayama K, Ohki O, et al. Comparative analysis of CD80 and CD86 on human Langerhans cells: expression and function. *Arch Dermatol Res.* 1998;290:547-552.
- 82. Wakem P Jr, Burns RP Jr, Ramirez F, et al. Allergens and irritants transcriptionally upregulate CD80 gene expression in human keratinocytes. *J Invest Dermatol*. 2000;114:1085-1092.
- 83. Fabbri M, Smart C, Pardi R. T lymphocytes. *Int J Biochem Cell Biol.* 2003;35:1004-1008.
- 84. Yawalkar N, Hari Y, Frutig K, et al. T cells isolated from positive epicutaneous test reactions to amoxicillin and ceftriaxone are drug specific and cytotoxic. *J Invest Dermatol*. 2000;115:647-652.
- 85. Naisbitt DJ, Britschgi M, Wong G, et al. Hypersensitivity reactions to carbamazepine: characterization of the specificity, phenotype, and cytokine profile of drug-specific T cell clones. *Mol Pharmacol*. 2003;63:732-741.
- 86. Schnyder B, Mauri-Hellweg D, Zanni M, et al. Direct, MHC-dependent presentation of the drug sulfamethoxazole to human alphabeta T cell clones. *J Clin Invest*. 1997;100:136-141.
- 87. Zanni MP, Mauri-Hellweg D, Brander C, et al. Characterization of lidocaine-specific T cells. *J Immunol*. 1997;158:1139-1148.
- 88. Zanni MP, von Greyerz S, Hari Y, et al. Recognition of local anesthetics by alphabeta+ T cells. *J Invest Dermatol*. 1999;112:197-204.
- 89. Gerber BO, Pichler WJ. Cellular mechanisms of T cell mediated drug hypersensitivity. *Curr Opin Immunol*. 2004;16:732-737.
- 90. Gerber BO, Pichler WJ. Noncovalent interactions of drugs with immune receptors may mediate drug-induced hypersensitivity reactions. *AAPS J.* In press.
- 91. Santamaria Babi LF, Perez Soler MT, Hauser C, et al. Skin-homing T cells in human cutaneous allergic inflammation. *Immunol Res.* 1995;14:317-324.
- 92. Santamaria LF, Perez Soler MT, Hauser C, et al. Allergen specificity and endothelial transmigration of T cells in allergic contact dermatitis and atopic dermatitis are associated with the cutaneous lymphocyte antigen. *Int Arch Allergy Immunol.* 1995;107:359-362.
- 93. Pichler WJ, Yawalkar N, Britschgi M, et al. Cellular and molecular pathophysiology of cutaneous drug reactions. *Am J Clin Dermatol*. 2002;3:229-238.

- 94. Nickoloff BJ, Turka LA. Immunological functions of nonprofessional antigen-presenting cells: new insights from studies of T-cell interactions with keratinocytes. *Immunol Today*. 1994;15:464-469.
- 95. Wikner NE, Huff JC, Norris DA, et al. Study of HLA-DR synthesis in cultured human keratinocytes. *J Invest Dermatol*. 1986:87:559-564.
- 96. Meunier L, Vian L, Lagoueyte C, et al. Quantification of CD1a, HLA-DR, and HLA class I expression on viable human Langerhans cells and keratinocytes. *Cytometry*. 1996;26:260-264.
- 97. Gueniche A, Viac J, Lizard G, et al. Effect of nickel on the activation state of normal human keratinocytes through interleukin 1 and intercellular adhesion molecule 1 expression. *Br J Dermatol*. 1994;131:250-256.
- 98. Gueniche A, Viac J, Lizard G, et al. Effect of various metals on intercellular adhesion molecule-1 expression and tumour necrosis factor alpha production by normal human keratinocytes. *Arch Dermatol Res.* 1994;286:466-470.
- 99. Dang LH, Michalek MT, Takei F, et al. Role of ICAM-1 in antigen presentation demonstrated by ICAM-1 defective mutants. *J Immunol*. 1990;144:4082-4091.
- 100. Piguet PF. Keratinocyte-derived tumor necrosis factor and the physiopathology of the skin. *Springer Semin Immunopathol*. 1992;13:345-354.
- 101. Terunuma A, Aiba S, Tagami H. Cytokine mRNA profiles in cultured human skin component cells exposed to various chemicals: a simulation model of epicutaneous stimuli induced by skin barrier perturbation in comparison with that due to exposure to haptens or irritant. *J Dermatol Sci.* 2001;26:85-93.
- 102. Ansel J, Perry P, Brown J, et al. Cytokine modulation of keratinocyte cytokines. *J Invest Dermatol*. 1990;94:101S-107S.
- 103. Sebastiani S, Albanesi C, De PO, et al. The role of chemokines in allergic contact dermatitis. *Arch Dermatol Res.* 2002;293:552-559.
- 104. Albanesi C, Scarponi C, Giustizieri ML, et al. Keratinocytes in inflammatory skin diseases. *Curr Drug Targets Inflamm Allergy*. 2005;4:329-334.
- 105. Cumberbatch M, Bhushan M, Dearman RJ, et al. IL-1beta-induced Langerhans' cell migration and TNF-alpha production in human skin: regulation by lactoferrin. *Clin Exp Immunol*. 2003;132:352-359.
- 106. Cumberbatch M, Dearman RJ, Griffiths CE, et al. Langerhans cell migration. *Clin Exp Dermatol*. 2000;25:413-418.
- 107. Cumberbatch M, Griffiths CE, Tucker SC, et al. Tumour necrosis factor-alpha induces Langerhans cell migration in humans. *Br J Dermatol.* 1999;141:192-200.
- 108. Nassif A, Moslehi H, Le Gouvello S, et al. Evaluation of the potential role of cytokines in toxic epidermal necrolysis. *J Invest Dermatol*. 2004;123:850-855.
- 109. Hulette BA, Ryan CA, Gerberick GF. Elucidating changes in surface marker expression of dendritic cells following chemical allergen treatment. *Toxicol Appl Pharmacol*. 2002;182:226-233.
- 110. Albanesi C, Cavani A, Girolomoni G. Interferon-gamma-stimulated human keratinocytes express the genes necessary for the production of peptide-loaded MHC class II molecules. *J Invest Dermatol*. 1998;110:138-142.
- 111. Basham TY, Nickoloff BJ, Merigan TC, et al. Recombinant gamma interferon induces HLA-DR expression on cultured human keratinocytes. *J Invest Dermatol*. 1984;83:88-90.

- 112. Lisby S, Muller KM, Jongeneel CV, et al. Nickel and skin irritants up-regulate tumor necrosis factor-alpha mRNA in keratinocytes by different but potentially synergistic mechanisms. *Int Immunol*. 1995;7:343-352.
- 113. Wilmer JL, Burleson FG, Kayama F, et al. Cytokine induction in human epidermal keratinocytes exposed to contact irritants and its relation to chemical-induced inflammation in mouse skin. *J Invest Dermatol.* 1994;102:915-922.
- 114. Piguet PF, Grau GE, Hauser C, et al. Tumor necrosis factor is a critical mediator in hapten induced irritant and contact hypersensitivity reactions. *J Exp Med*. 1991;173:673-679.
- 115. Vandebriel RJ, Van Och FM, van Loveren H. In vitro assessment of sensitizing activity of low molecular weight compounds. *Toxicol Appl Pharmacol*. 2005;207:142-148.
- 116. O'Garra A, McEvoy LM, Zlotnik A. T-cell subsets: chemokine receptors guide the way. *Curr Biol*. 1998;8:R646-R649.
- 117. Lebrec H, Kerdine S, Gaspard I, et al. Th(1)/Th(2) responses to drugs. *Toxicology*. 2001;158:25-29.
- 118. Umetsu DT, DeKruyff RH. Th1 and Th2 CD4+ cells in the pathogenesis of allergic diseases. *Proc Soc Exp Biol Med*. 1997;215:11-20.
- 119. Xu H, DiIulio NA, Fairchild RL. T cell populations primed by hapten sensitization in contact sensitivity are distinguished by polarized patterns of cytokine production: interferon gamma-producing (Tc1) effector CD8+ T cells and interleukin (Il) 4/Il-10-producing (Th2) negative regulatory CD4+ T cells. *J Exp Med*. 1996;183:1001-1012.
- 120. Fuchs J, Zollner TM, Kaufmann R, et al. Redox-modulated pathways in inflammatory skin diseases. *Free Radic Biol Med*. 2001;30:337-353.
- 121. Steinbrink K, Sorg C, Macher E. Low zone tolerance to contact allergens in mice: a functional role for CD8+ T helper type 2 cells. *J Exp Med.* 1996;183:759-768.
- 122. Dearman RJ, Basketter DA, Kimber I. Characterization of chemical allergens as a function of divergent cytokine secretion profiles induced in mice. *Toxicol Appl Pharmacol*. 1996;138:308-316.
- 123. Dieli F, Asherson GL, Sireci G, et al. Development of IFN-gamma-producing CD8+ gamma delta+ T lymphocytes and IL-2-producing CD4+ alpha beta+ T lymphocytes during contact sensitivity. *J Immunol*. 1997;158:2567-2575.
- 124. Blaise GA, Gauvin D, Gangal M, et al. Nitric oxide, cell signaling and cell death. *Toxicology*. 2005;208:177-192.
- 125. Ross R, Reske-Kunz AB. The role of NO in contact hypersensitivity. *Int Immunopharmacol*. 2001;1:1469-1478.
- 126. Bruch-Gerharz D, Ruzicka T, Kolb-Bachofen V. Nitric oxide in human skin: current status and future prospects. *J Invest Dermatol*. 1998:110:1-7.
- 127. Weller R. Nitric oxide: a key mediator in cutaneous physiology. *Clin Exp Dermatol.* 2003;28:511-514.
- 128. Deliconstantinos G, Villiotou V, Stravrides JC. Release by ultraviolet B (u.v.B) radiation of nitric oxide (NO) from human keratinocytes: a potential role for nitric oxide in erythema production. *Br J Pharmacol.* 1995;114:1257-1265.
- 129. Nathan C. Inducible nitric oxide synthase: what difference does it make? *J Clin Invest*. 1997;100:2417-2423.
- 130. Arany I, Brysk MM, Brysk H, et al. Induction of iNOS mRNA by interferon-gamma in epithelial cells is associated with growth arrest and differentiation. *Cancer Lett.* 1996;110:93-96.

- 131. Qureshi AA, Hosoi J, Xu S, et al. Langerhans cells express inducible nitric oxide synthase and produce nitric oxide. *J Invest Dermatol*. 1996;107:815-821.
- 132. Rocha IM, Guillo LA. Lipopolysaccharide and cytokines induce nitric oxide synthase and produce nitric oxide in cultured normal human melanocytes. *Arch Dermatol Res.* 2001;293:245-248.
- 133. Chang HR, Tsao DA, Wang SR, et al. Expression of nitric oxide synthases in keratinocytes after UVB irradiation. *Arch Dermatol Res.* 2003;295:293-296.
- 134. Warren JB. Nitric oxide and human skin blood flow responses to acetylcholine and ultraviolet light. *FASEB J.* 1994;8:247-251.
- 135. Lerner LH, Qureshi AA, Reddy BV, et al. Nitric oxide synthase in toxic epidermal necrolysis and Stevens-Johnson syndrome. *J Invest Dermatol.* 2000;114:196-199.
- 136. Rowe A, Farrell AM, Bunker CB. Constitutive endothelial and inducible nitric oxide synthase in inflammatory dermatoses. *Br J Dermatol.* 1997;136:18-23.
- 137. Lippe IT, Stabentheiner A, Holzer P. Participation of nitric oxide in the mustard oil-induced neurogenic inflammation of the rat paw skin. *Eur J Pharmacol*. 1993;232:113-120.
- 138. Morita H, Hori M, Kitano Y. Modulation of picryl chloride-induced contact hypersensitivity reaction in mice by nitric oxide. *J Invest Dermatol.* 1996;107:549-552.
- 139. Ross R, Gillitzer C, Kleinz R, et al. Involvement of NO in contact hypersensitivity. *Int Immunol*. 1998;10:61-69.
- 140. Nathan C, Xie QW. Nitric oxide synthases: roles, tolls, and controls. *Cell*. 1994;78:915-918.
- 141. Forstermann U, Closs EI, Pollock JS, et al. Nitric oxide synthase isozymes: characterization, purification, molecular cloning, and functions. *Hypertension*. 1994;23:1121-1131.
- 142. Wanikiat P, Woodward DF, Armstrong RA. Investigation of the role of nitric oxide and cyclic GMP in both the activation and inhibition of human neutrophils. *Br J Pharmacol*. 1997;122:1135-1145.
- 143. Shin WS, Hong YH, Peng HB, et al. Nitric oxide attenuates vascular smooth muscle cell activation by interferon-gamma: the role of constitutive NF-kappa B activity. *J Biol Chem.* 1996;271:11317-11324.
- 144. Bonham CA, Lu L, Li Y, et al. Nitric oxide production by mouse bone marrow-derived dendritic cells: implications for the regulation of allogeneic T cell responses. *Transplantation*. 1996;62:1871-1877.
- 145. Lu L, Bonham CA, Chambers FG, et al. Induction of nitric oxide synthase in mouse dendritic cells by IFN-gamma, endotoxin, and interaction with allogeneic T cells: nitric oxide production is associated with dendritic cell apoptosis. *J Immunol*. 1996;157:3577-3586.
- 146. Virag L, Szabo E, Bakondi E, et al. Nitric oxide-peroxynitrite-poly(ADP-ribose) polymerase pathway in the skin. *Exp Dermatol*. 2002;11:189-202.
- 147. Briganti S, Picardo M. Antioxidant activity, lipid peroxidation and skin diseases: what's new. *J Eur Acad Dermatol Venereol*. 2003:17:663-669.
- 148. Chain BM. Current issues in antigen presentation: focus on the dendritic cell. *Immunol Lett.* 2003;89:237-241.
- 149. Hubbard AK, Rothlein R. Intercellular adhesion molecule-1 (ICAM-1) expression and cell signaling cascades. *Free Radic Biol Med.* 2000;28:1379-1386.
- 150. Rutault K, Alderman C, Chain BM, et al. Reactive oxygen species activate human peripheral blood dendritic cells. *Free Radic Biol Med.* 1999;26:232-238.

- 151. Verhasselt V, Goldman M, Willems F. Oxidative stress up-regulates IL-8 and TNF-alpha synthesis by human dendritic cells. *Eur J Immunol*. 1998;28:3886-3890.
- 152. Mates JM, Perez-Gomez C, Olalla L, et al. Allergy to drugs: antioxidant enzymic activities, lipid peroxidation and protein oxidative damage in human blood. *Cell Biochem Funct*. 2000;18:77-84.
- 153. Coopman SA, Johnson RA, Platt R, et al. Cutaneous disease and drug reactions in HIV infection. *N Engl J Med.* 1993;328:1670-1674.
- 154. Buhl R, Jaffe HA, Holroyd KJ, et al. Systemic glutathione deficiency in symptom-free HIV-seropositive individuals. *Lancet*. 1989;2:1294-1298.
- 155. Kaur S, Zilmer M, Eisen M, et al. Patients with allergic and irritant contact dermatitis are characterized by striking change of iron and oxidized glutathione status in nonlesional area of the skin. *J Invest Dermatol.* 2001;116:886-890.
- 156. Kaur S, Zilmer M, Eisen M, et al. Nickel sulphate and epoxy resin: differences in iron status and glutathione redox ration at the time of patch testing. *Arch Dermatol Res.* 2004;295:517-520.
- 157. Nordberg J, Zhong L, Holmgren A, et al. Mammalian thioredoxin reductase is irreversibly inhibited by dinitrohalobenzenes by alkylation of both the redox active selenocysteine and its neighboring cysteine residue. *J Biol Chem.* 1998;273:10835-10842.
- 158. Vyas PM, Roychowdhury S, Woster PM, et al. Reactive oxygen species generation and its role in the differential cytotoxicity of the arylhydroxylamine metabolites of sulfamethoxazole and dapsone in normal human epidermal keratinocytes. *Biochem Pharmacol*. 2005;70:275-286.
- 159. Kantengwa S, Jornot L, Devenoges C, et al. Superoxide anions induce the maturation of human dendritic cells. *Am J Respir Crit Care Med*. 2003;167:431-437.
- 160. Chen KH, Reece LM, Leary JF. Mitochondrial glutathione modulates TNF-alpha-induced endothelial cell dysfunction. *Free Radic Biol Med.* 1999;27:100-109.

- 161. Ikeda M, Schroeder KK, Mosher LB, et al. Suppressive effect of antioxidants on intercellular adhesion molecule-1 (ICAM-1) expression in human epidermal keratinocytes. *J Invest Dermatol*. 1994;103:791-796.
- 162. Faruqi RM, Poptic EJ, Faruqi TR, et al. Distinct mechanisms for N-acetylcysteine inhibition of cytokine-induced E-selectin and VCAM-1 expression. *Am J Physiol*. 1997;273:H817-H826.
- 163. Matsue H, Edelbaum D, Shalhevet D, et al. Generation and function of reactive oxygen species in dendritic cells during antigen presentation. *J Immunol*. 2003;171:3010-3018.
- 164. Sandstrom PA, Buttke TM. Autocrine production of extracellular catalase prevents apoptosis of the human CEM T-cell line in serum-free medium. *Proc Natl Acad Sci USA*. 1993;90:4708-4712.
- 165. Kannan K, Jain SK. Oxidative stress and apoptosis. *Pathophysiology*. 2000;7:153-163.
- 166. Ogawa Y, Kobayashi T, Nishioka A, et al. Reactive oxygen species-producing site in hydrogen peroxide-induced apoptosis of human peripheral T cells: involvement of lysosomal membrane destabilization. *Int J Mol Med.* 2004;13:383-388.
- 167. Devadas S, Zaritskaya L, Rhee SG, et al. Discrete generation of superoxide and hydrogen peroxide by T cell receptor stimulation: selective regulation of mitogen-activated protein kinase activation and fas ligand expression. *J Exp Med.* 2002;195:59-70.
- 168. Hildeman DA, Zhu Y, Mitchell TC, et al. Activated T cell death in vivo mediated by proapoptotic bcl-2 family member bim. *Immunity*. 2002;16:759-767.
- 169. Fuchs J, Packer L. Antioxidant protection from solar-simulated radiation-induced suppression of contact hypersensitivity to the recall antigen nickel sulfate in human skin. *Free Radic Biol Med.* 1999;27:422-427.
- 170. Pasche-Koo F, Arechalde A, Arrighi JF, et al. Effect of N-acetylcysteine, an inhibitor of tumor necrosis factor, on irritant contact dermatitis in the human. *Curr Probl Dermatol.* 1995;23:198-206.